

## A NEW POTENTIOMETRIC SENSOR FOR THE ASSAY OF P53 IN BLOOD SAMPLES

Ahmed Jassim Muklive AL-OGAIDI<sup>1</sup>, Raluca-Ioana STEFAN-VAN STADEN<sup>2</sup>,  
Livia Alexandra GUGOASA<sup>3</sup>, Jacobus Frederick VAN STADEN<sup>4</sup>, Hülya  
YANIK<sup>5</sup>, Meltem GÖKSEL<sup>6</sup>, Mahmut DURMUŞ<sup>7</sup>

*A potentiometric sensor based on graphite paste modified with a phthalocyanine-BODIPY conjugate was proposed for the assay of tumor suppressor P53. The working concentration range was between  $1.17 \times 10^{-9}$  and  $2.93 \times 10^{-8} \mu\text{g/mL}$  with a slope of  $82.62 \text{ mV}/\mu\text{g mL}^{-1}$  and a limit of detection was  $3.37 \times 10^{-10} \mu\text{g/mL}$ . The selectivity of the sensor was checked against possible interfering species from blood samples. The sensor was applied for the analysis of P53 in whole blood samples, without any pretreatment of the sample.*

**Keywords:** P53, colon cancer, potentiometric sensor, blood samples.

### List of Abbreviations

MWCNT- multiwalled carbon nanotube  
PA6 – doped nylon 6  
PPy – polypyrrole  
GCE –glassy carbon electrode  
t-GO – thiolated graphene oxide

<sup>1</sup> Ph.D Student., Faculty of Applied Chemistry and Materials Science, University POLITEHNICA of Bucharest, Romania, e-mail: ahmed.mukleef@yahoo.com

<sup>2</sup> Prof. Dr. Habil, Department of Analytical Chemistry and Environmental Engineering, Faculty of Applied Chemistry and Materials Science, University POLITEHNICA of Bucharest; Laboratory of Electrochemistry and PATLAB Bucharest, National Institute of Research for Electrochemistry and Condensed Matter, Bucharest, Romania e-mail: ralucavanstaden@gmail.com

<sup>3</sup> Researcher, PhD, Laboratory of Electrochemistry and PATLAB Bucharest, National Institute of Research for Electrochemistry and Condensed Matter, Bucharest, Romania e-mail: livia.gugoasa@yahoo.com

<sup>4</sup> Professor Dr, Laboratory of Electrochemistry and PATLAB Bucharest, National Institute of Research for Electrochemistry and Condensed Matter, Bucharest, Romania e-mail: koosvanstaden2012@yahoo.com

<sup>5</sup> Researcher, PhD, Department of Chemistry, Gebze Technical University, Gebze-Kocaeli, Turkey, e-mail: hulyayanik82@hotmail.com

<sup>6</sup> Assistant Professor Dr, Kocaeli University, Kosekoy Vocational School, Department of Chemistry, Kartep, Kocaeli, Turkey, e-mail: meltemgoksel@gmail.com

<sup>7</sup> Professor Dr, Department of Chemistry, Gebze Technical University, Gebze-Kocaeli, Turkey, e-mail: durmus@gtu.edu.tr

bt - biotinilated  
 NPs – nanoparticles  
 BSA – bovine serum albumin  
 ABC – avidin-biotin-peroxidase complex  
 BODIPY - boron dipyrromethene  
 ECL - Electrochemiluminescence  
 DPV - Differential pulse voltammetry  
 IM-ECL - Immunomagnetic-electrochemiluminescence

## 1. Introduction

P53 is a well-known tumor suppressor gene that plays a vital role in the repairing of DNA, apoptosis and cell proliferation. It conserves the stability in cells by preventing genome mutation [1]. Moreover, the deactivation of p53 gene is the most commonly detected anomaly in human cancers and was noticed in more than half of the human cancers [2]. The cell proliferation is disrupted and tumors are formed once P53 is mutated. Also, p53 gene is mutated in 50% of human tumors [3]. Most studies have stated that mutant P53 is highly expressed and accumulated in cancer cells due to its prolonged half-life [4]. Furthermore, P53 mutation in serum could be a useful marker for the diagnosis of human colon cancer [5]. Consequently, the pathway of P53 plays a major role in regulation of cell growth and survival, in which the detection of P53 mutant level in human serum is an effective way for early diagnosis and prognosis of cancer. Numerous research works have been conducted regarding the quantitative detection of P53 for this purpose.

Several sensing techniques have been developed for detection of P53 such as surface plasmon resonance (SPR) [6], field-effect transistors [7], quartz crystal microbalance [8], colorimetry [9], chemiluminescence [10], enzyme-linked immunosorbent assay (ELISA) [11], and different electrochemical methods which are presented in Table 1. In this paper we proposed a potentiometric sensor based on graphite paste modified with a phthalocyanine-BODIPY conjugate (Figure 1) for the assay of P53 in blood samples.

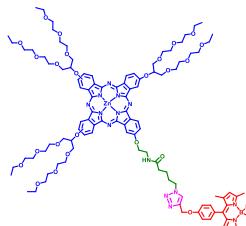


Fig. 1. Chemical structure of phthalocyanine-Bodipy conjugate

Table 1

**Different methods for the analysis of P53 tumor suppressor.**

Method	Electrode type	LOD	LOQ	Linear concentration range	Biological sample	Ref.
DPV- <i>biosensor</i>	MWCNT-PA6-PPy/Au	50 fmol/L	100 fmol/L	0.1 pmol/L – 100 pmol/L	-	12
ECL <i>ECL immunosensor</i>	Ru–silica nanoporous@gold nanocomposite/GCE	22.8 fmol/L	76.1 fmol/L	0.2 pmol/L - 200 pmol/L	Serum Skin fibroblast cells	13
SPR <i>SPR chip</i>	Carboxymethylated-dextran modified Au sensor chip	1.06 pmol/L	0.01 nmol/L	0.01 nmol/L – 1.06 nmol/L	Normal/ Cancer cells	14
DPV <i>immunosensor</i>	t-GO/Streptavidin-Au NPs/bt antibody /BSA/antigen/bt secondary antibody/ABC/GCE	30 fmol/L	115 fmol/L	0.2 pmol/L - 200 pmol/L	Serum Skin fibroblast cells	15
(IM-ECL)	Pt Electrode	-	0.01 ng/mL	0.01 ng/mL - 1000 ng/mL	Serum	16
Potentiometry <i>potentiometric sensor</i>	Phthalocyanine-boron dipyrromethene /Graphite	0.34 fg/mL	1.17 fg/mL	1.17 fg/mL – 29.3 fg/mL	Whole blood	This work

**2. Experimental****2.1. Reagents and materials**

Tumor suppressor P53, graphite powder, monosodium phosphate and disodium phosphate were purchased from Sigma Aldrich (Milwaukee, USA). MEG107porphyrin ( $C_{106}H_{139}BF_2N_{14}O_{24}Zn$ ) was synthesized in the house by the group of Professor Mahmut Durmus. Paraffin oil ( $d_4^{20}$ , 0.86 g x  $cm^{-1}$ ) was purchased from Fluka (Buchs, Switzerland). Monosodium phosphate and disodium phosphate were used for preparation of phosphate buffer 0.1 mol/L, pH=7.5. Deionized water obtained from a Millipore Direct-Q3 System (Molsheim, France) was used for the preparation of all solution. Standard solution of P53 was prepared in PBS pH=7.5 to a concentration of 7.14 $\mu$ g/mL. Serial dilution technique was used for the preparation of solutions of different concentration for P53. All solutions were of analytical grade.

**2.2. Apparatus**

For all potentiometric measurements an Ivium potentiostat/galvanostat was used. The potentiometric cell is composed from a two-electrode system: reference and working electrode. Ag/AgCl electrode served as reference electrode in the cell.

### 2.3. Sensor's design

Modified graphite paste was prepared as follows: 100 mg of graphite powder was mixed with 30  $\mu$ L of paraffin oil to form a homogenous paste. 100  $\mu$ L of  $10^{-3}$  mol L $^{-1}$  phthalocyanine-BODIPY conjugate was added to this paste, to obtain the modified paste. Electric contact was made using a silver wire (0.5 mm in diameter). Between the measurements, the sensor was washed with deionized water. When not in use, the graphite potentiometric sensor was kept in a dry place at room temperature.

### 2.4. Recommended procedure

Direct potentiometry was used for the measurements of the potential of each solution of known concentration (1.17 fg/mL – 7.14  $\mu$ g/mL). The working electrode was placed in stirred standard solution while the potential was recorded. The equation of calibration was obtained using statistics, and used for calculations of unknown concentrations of P53 in whole blood samples.

### 2.5. Sample preparation for whole blood

Whole blood samples were obtained from the University Hospital in Bucharest (Ethics committee approval nr. 75/2015) from 5 different patients diagnosed with colon cancer. These samples were used for the assay of P53 without any pretreatment. The cell was filled with the whole blood and the potential was measured for 5 minutes for each sample. The unknown concentrations were determined from the calibration curve as described in the direct potentiometry method.

## 3. Results and discussions

### 3.1. Characteristic response of potentiometric sensors

The mechanism of P53 recognition by phthalocyanine BODIPY is based on formation of host-guest compound between the analyte (P53) and ligand (phthalocyanine BODIPY) at the membrane-solution interface [17].

The sensor based on graphite paste modified with phthalocyanine-BODIPY conjugate was tested using direct potentiometric method for the analysis of P53. The mechanism of potential development was explained by the interaction between P53 and phthalocyanine-BODIPY conjugate at the membrane-solution interface. The proposed sensor showed linear and near-Nernstian response, so that it can be used for the determination of P53. The sensor based on graphite paste modified with phthalocyanine-BODIPY conjugate can be used for the assay of P53 in the linear concentration range 1.17 fg/mL – 29.3 fg/mL. The response time of the sensor was 5 minutes. The limit of quantification reached with the sensor based on phthalocyanine-BODIPY conjugate/graphite for P53 was 1.17 fg/mL. The proposed sensor covered the range on which P53 can be found on patients not

clinical ill as well as for patients presenting stages 1 - 3 of colon cancer. The equation of calibration recorded was:

$$E = -617.4 + 82.6 \times pP53; r = 0.9999 \quad (1)$$

where  $pP53$  is  $-\log C_{P53}$  and  $r$  is the correlation coefficient. The standard potential was  $-617.4 \pm 2.47$  mV, and the slope of the sensor was  $82.6 \pm 0.93$  mV/decade of concentration. The limit of detection was 0.34 fg/mL. Calibration graph is shown in Figure 2.

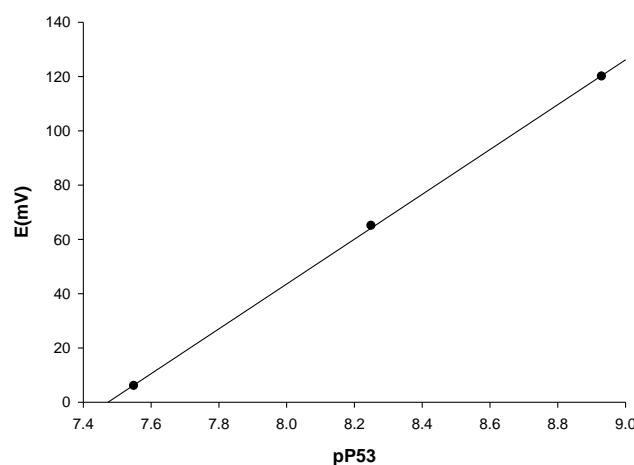


Fig. 2. Calibration graph of the potentiometric sensor

The sensor was used for more than one month for measurements, and calibrated every day. The RSD of the slope of the proposed sensor was 1.25% after 30 days of daily use. Also, 10 such pastes were made and used in sensors' bodies for measurements – when the values of RSD of the slopes obtained was 0.24%, showing good reproducibility of sensor's design.

The response characteristics of the proposed sensor made possible the detection of P53 in whole blood at a very early stage, when its concentration is less than  $10 \text{ ng mL}^{-1}$  [12-16].

The advantages of the proposed sensor and method for the determination of P53 are versus the other methods proposed to date [12-16]: it has the lowest determination and detection limits; the range of the working concentration favorized its early detection in the biological samples; it is the only one that was used for whole blood screening.

### 3.2. Selectivity

Dopamine (DA), norepinephrine (NE), epinephrine (Epi), acetylcholine (Ach), kirsten rat sarcoma viral (KRAS), human epidermal growth factor receptor

1 (HER1) and carcinoembryonic (CEA) have been chosen as possible interfering species in the detection of P53. Mixed solution method was employed to determine the potentiometric selectivity coefficient. The ratio between the concentration (mol/L) of the interfering species and P53 was 10:1. The results are shown in Table 2. The values obtained for potentiometric selectivity coefficients shown that the selected substances are slightly interfering with the determination of P53.

*Table 2*  
**Potentiometric selectivity coefficients for the sensor based on phthalocyanine-BODIPY conjugate/Graphite**

Potentiometric sensor based on	$K_{sel}^{pot}$						
	DA	Epi	NE	Ach	KRAS	HER-1	CEA
Phthalocyanine-BODIPY conjugate/Graphite	$2.1 \times 10^{-3}$	$1.2 \times 10^{-3}$	$1.2 \times 10^{-3}$	$1.8 \times 10^{-3}$	$1.5 \times 10^{-3}$	$1.5 \times 10^{-3}$	$7.7 \times 10^{-3}$

Taking into account the mechanism of potential development specific for potentiometric sensors, the main explanation for high selectivity of the proposed sensor versus selected possible interferences is that the ligand is forming less stable complex compared with those formed by P53.

### 3.3. Analytical applications

The characteristics of response recorded for the sensor based on phthalocyanine-BODIPY conjugate/Graphite (linear concentration range of determination, selectivity, and sensitivity) revealed that the sensor could be used for the evaluation of P53 in biological fluids. Consequently, the proposed sensor was used for the analysis of P53 in whole blood samples. The five samples of blood were supplied from the hospital, and used as collected, for the evaluation of P53 by using the proposed sensor. Therefore, the results obtained by using the sensor based on phthalocyanine-BODIPY conjugate/Graphite show that it is a reliable tool for the evaluation of P53 in whole blood samples (Table 3). The proposed sensor based on phthalocyanine-BODIPY conjugate/Graphite can detect P53 at lower concentrations.

*Table 3*  
**Recovery tests of P53 in blood samples.**

Sample no.	fg/mL, Recovery of P53				
	1	2	3	4	5
	$5.62 \pm 0.07$	$6.76 \pm 0.09$	$5.00 \pm 0.07$	$4.68 \pm 0.08$	$1.17 \pm 0.08$

A control sample taken from a healthy patient showed no P53 in the blood sample.

#### 4. Conclusions

The proposed sensor shown very good results for the recovery test which makes it a reliable tool for measuring P53 in whole blood samples, and this is very important for early detection of colon cancer. The proposed method was highly sensitive, exhibited good selectivity for the detection of P53 from blood samples. The advantages of the proposed method versus techniques like ELISA and chromatography are the following: it is a simple and easy method performed with low cost, short analysis time, and low limit of quantification (1.17 fg/mL), being able to detect at an early stage the colon cancer.

#### Acknowledgements

This work was supported by PNII Program Partnership 2014–2016, MULTIMODESCREEN, Contract no. 22/2014.

#### R E F E R E N C E S

- [1]. *C. Prives, P. A. Hall*, “The p53 pathway”, *The Journal of Pathology*, **vol. 187**, 1999, pp. 112-126.
- [2]. *O.3 Onen, A. Sisman, N. D. Gallant, P. Kruk and R. Guldiken*, “A Urinary Bcl-2 Surface Acoustic Wave Biosensor for Early Ovarian Cancer Detection”, *Sensors*, **vol. 12**, 2012, pp. 7423-7437.
- [3]. *H. Bouzourene, P. Gervaz, J.-P. Cerottini, J. Benhattar, P. Chaubert, E. Saraga, S. Pampallona, F.T. Bosman and J.-C. Givel*, “p53 and Ki-ras as prognostic factors for Dukes' stage B colorectal cancer”, *European journal of Cancer*, **vol. 36**, 2000, pp.1008-1015.
- [4]. *S. Strano, S. Dell'Orso, S.D. Agostino, G. Fontemaggi, A. Sacchi, G. Blandino*, “Mutant p53: an oncogenic transcription factor”, *Oncogene*, **vol. 26**, 2007, pp. 2212-2215.
- [5]. *M.C. Hulvat, J. S. Jeruss* “Maintaining Fertility in Young Women with Breast Cancer” *Current Treatment Options in Oncology*, **vol. 10**, 2009, pp.308-317.
- [6]. *T. Jiang, M. Minunni, P. Wilson, J. Zhang and A.P.F. Turner, M. Mascini* “Detection of TP53 mutation using a portable Surface Plasmon Resonance DNA-based Biosensor”, *Biosensors and Bioelectronics*, **vol. 20**, 2005, pp.1939-1945.
- [7]. *S.H. Han, S.K. Kim, K. Park, S.Y. Yi, H. J. Park, H.K. Lyu, M. Kim and B.H. Chung*, “Detection of mutant p53 using field-effect transistor biosensor”, *Analytica Chemica Acta*, **vol. 665**, 2010, pp. 79-83.
- [8]. *Z. Altintas, I.E. Tothill*, “DNA-based biosensor platforms for the detection of TP53mutation”, *Sensors and Actuators B: Chemical*, **vol. 169**, 2012, pp. 188-194.
- [9]. *H. Li, Z. Wu, L. Qiu, J. Liu, C. Wang, G. Shen and R. Yu*, “Ultrasensitive label-free amplified colorimetric detection of p53 based on G-quadruplex MBzymes”, *Biosensors and Bioelectronics*, **vol. 50**, 2013, pp.180-185.
- [10]. *X. Chen, C. He, Z. Zhang and J. Wang*, “Sensitive chemiluminescence detection of wild-type p53 protein captured by surface-confined consensus DNA duplexes”, *Biosensors and Bioelectronics*, **vol. 47**, 2013, pp. 335-339.
- [11]. *J. M. Portefaix, Fanutti, C. Granier, C. Crapez, E. Perham, R. Grenier, J. Pau and B. Del Rio*, “Detection of anti-p53 antibodies by ELISA using p53 synthetic or phage-displayed peptides”, *Journal of Immunological Methods*, **vol. 259**, 2002, pp. 65-75.

- [12]. *A. Afsharan, F. Navaeipour, B. Khalizadeh, H. Tajalli, M. Mollabashi, M.J. Ahar, M.R. Rashidi*, “Highly sensitive electrochemiluminescence detection of p53 protein using functionalized Ru-silica nanoporous@gold nanocomposite”, *Biosensors and Bioelectronics.*, **vol. 80**, 2016, pp. 146-153.
- [13]. *G. Yan, D. Xing, S. Tan, Q. Chen*, “Rapid and sensitive immunomagnetic electrochemiluminescent detection of p53 antibodies in human serum”, *Journal of Immunological Methods.*, **vol. 288**, 2004, pp. 47-54.
- [14] *H. Afsharan, B. Khalilzahed, H. Tajalli, M. Mollabashi, F. Navaeipour, M.R. Rashidi*, “A sandwich type immunosensor for ultrasensitive electrochemical quantification of p53 protein based on gold nanoparticles/graphene oxide”, *Electrochimica Acta.*, **vol. 188**, 2016, pp. 153-164.
- [15]. *X. Wang, X. Wang, X. Wang, F. Chen, K. Zhu, Q. Xu, M. Tang*, “Novel electrochemical biosensor based on functional composite nanofibers for sensitive detection of p53 tumor suppressor gene”, *Analytica Chimica Acta.*, **vol. 765**, 2013, pp. 63-69.
- [16]. *Y. Wang, X. Zhu, M. Wu, N. Xia, J. Wang, F. Zhou*, “Simultaneous and label-free determination of wild – type and mutant p53 at a single surface Plasmon resonance chip preimmobilized with consensus DNA and Monoclonal Antibody”, *Analytical Chemistry*, **Vol. 81**, 2009, pp. 8441-8446.
- [17]. *R.I. Stefan, J.F. van Staden, H.Y. Aboul-Enein*, “Electrochemical Sensors in Bioanalysis”, 2001, Marcel Dekker Inc., New York, USA.